

# Two Automatic Chamber Techniques for Measuring Soil-Atmosphere Exchanges of Trace Gases and Results of their use in the OASIS Field Experiment

C.P. (Mick) Meyer, Ian E. Galbally, Yingping Wang, Ian A. Weeks  
*CSIRO Atmospheric Research*

and

Ian Jamie, David W.T. Griffith  
*Chemistry Department, University of Wollongong*



**Atmospheric Research**

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*CSIRO Atmospheric Research***

**Jamie, I. and Griffith, D.W.T.  
*Chemistry Department, University of Wollongong***

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Address and contact details: CSIRO Atmospheric Research  
Private Bag No. 1., Aspendale Victoria 3195 Australia  
Ph: (+61 3) 9239 4400; fax: (+61 3) 9239 4444  
email: [chief@dar.csiro.au](mailto:chief@dar.csiro.au)

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**TWO AUTOMATIC CHAMBER TECHNIQUES FOR MEASURING  
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C.P. (Mick) Meyer, Ian E. Galbally, Yingping Wang,  
Ian A. Weeks

*CSIRO Atmospheric Research  
Aspendale, Victoria, Australia*

and

Ian Jamie, David W.T. Griffith  
*Chemistry Department, University of Wollongong  
Wollongong, New South Wales, Australia*

**Abstract**

Two techniques for measuring soil/plant/atmosphere exchange fluxes of stable trace gases are presented. The first technique involves an automatic chamber capable of running at remote, unpowered, unattended sites. The air samples collected from individual flux measurements are progressively added to holding volumes, essentially averaging the concentrations in the accumulated air samples before their chemical analysis. This technique allows the continuous measurement of trace gas exchange at multiple distant remote sites to be carried out by one or two persons operating from a central laboratory housing the analytical facilities. The scientific advantages of this method are that emission measurements are automated, integrated day and night, and can be carried out at remote locations without mains power. The second technique involves continuous online analysis of the chamber headspace once per minute by Fourier Transform Infrared (FTIR) spectrometry. This technique provides simultaneous measurement of CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub>, is suitable for use at well supported measurement sites, and allows the continuous, automated determination of trace gas exchange measurements on a half hourly or hourly basis. The underlying theoretical and experimental basis of the use of chambers to measure trace gas exchange over soil/plant surfaces is presented. Measurements are presented from the OASIS (Observations At Several Interacting Scales), field experiment. The temperature and soil moisture responses of soil/plant respiration, N<sub>2</sub>O emissions and CH<sub>4</sub> uptake are observed and quantified. Both approaches produced comparable results for CO<sub>2</sub> fluxes. While for the most part N<sub>2</sub>O fluxes were also equivalent, there were some differences due principally to minor differences in chamber operating conditions.

**1. Introduction**

Analysis of chamber measurements is based on the application of the equation of conservation of mass for the trace gas within the chamber. Chamber measurements are particularly useful for studies of the soil and plant processes involved in this trace gas exchange. The particular requirements for accurate chamber measurements (Galbally and Roy, 1980; Galbally et al., 1985; Businger, 1986) are:

- a realistic and quantified boundary layer resistance within the chamber,
- adequate turbulent mixing of the free volume within the chamber,
- the inertness of the material making up the chamber wall,
- the equality of ambient pressure within and outside the chamber,
- minimal interference of the chamber with key soil and plant environmental variables; and
- a quantified rate of flow of air into and out of the chamber.

In the OASIS project (Leuning et al., 1997), measurements of trace gas exchange were made with different systems that measured on scales varying from a leaf to a landscape scale. This paper presents details about the chamber technique that makes individual measurements on patches of  $< 1 \text{ m}^2$  at multiple sites in a field. Two differently configured systems for measurement of the trace gas concentrations have been used with these chambers. One is a bag-sampling system for integrated flux measurements suitable for use at remote locations in the absence of electrical power, the other is a continuous system for use at well-supported field sites. Examples of the system performance are drawn from the OASIS fieldwork along with previous studies using this instrumentation.

## 2. Equipment and site description

### 2.1 Automatic flux chamber design

Each automatic flux chamber used in this experiment consisted of an open base unit that was pressed into the soil to a depth of 75 to 100 mm and a moveable box. The moveable boxes were constructed from anodised aluminium, have a volume of ca. 100 L and cover a ground area of  $0.35 \text{ m}^2$  (0.7 m by 0.5 m). The roof of each chamber was fitted with a glass window to maintain some natural lighting, along with a lid that closes or opens with a double-acting gas piston using compressed air from a small gas cylinder. The box unit fitted over the base and sealed with a silicone rubber O-ring compressed between two flanges (Fig. 1).

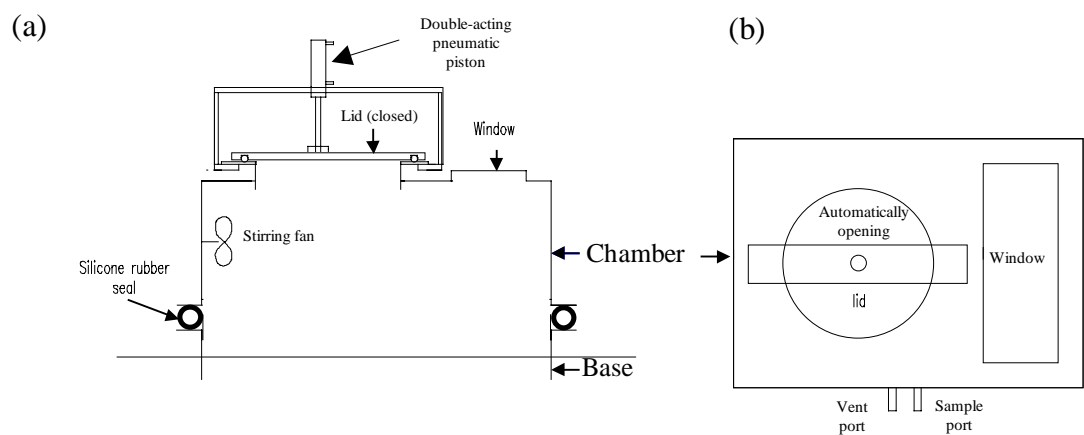


Figure 1 Schematic diagram of the chamber in (a) elevation and (b) plan.

Air within the chamber was continually stirred with an instrument fan to maintain a boundary-layer conductance determined as approximately  $0.02 \text{ m s}^{-1}$ . Pressure within the chamber was maintained at ambient at all times by a vent consisting of a 3-mm bulkhead union and 100-mm Teflon tube of internal diameter 2 mm. The operation of the chambers was controlled by a programmable microprocessor and all operations (i.e. digital output states, battery voltage, sample manifold pressure) and key environmental variables including chamber temperature, ambient (external) windspeed, and wind direction were logged at 30-s intervals.

When measurements were made over crops that are taller than the height of the standard chamber (0.3 m), an extension was fitted to the sides of the chambers doubling or tripling its height.

In the integrated method, air samples were drawn from the chamber by a 12 V Teflon pump (Model MPU487-NO5, KNF Neuberger, NJ, USA) powered by a regulated 12 V DC/DC converter to ensure a constant flow rate. The sample stream was then directed via one of five Teflon valves (Biovalve 1101461, Furon, USA) to either one of four Tedlar sample bags (8 litre or 11 litre, Cole Parmer, USA) or to the sample vent.

In the continuous method, air was circulated at  $15 \text{ L min}^{-1}$  through a closed loop from the chamber through approximately 30 m of 12-mm copper and Teflon tubing to the measurement cell in the FTIR spectrometer, and returned via an oil-free diaphragm pump (Thomas OD5) to the chamber.

The controller/logger and gas sampling equipment were housed in a weather-proof field housing. The logger/controller was based on an Intel 80552 microprocessor. The entire system was powered by a 12 V, 60 ampere-hour deep-cycle battery, which stored sufficient charge for 4 days' operation. When in automatic mode, the microprocessor programme accessed an event table specifying the required sequence of timed events. Communication with the logger/controller was via an RS232 interface to an ASCII terminal, usually a laptop PC, which was connected to the logger/controller to download the data during a daily maintenance visit to the site.

## **2.2 Operation and gas analysis**

### *2.2.1 Integrating method - gas chromatography*

The concentrations of  $\text{N}_2\text{O}$ ,  $\text{CH}_4$  and  $\text{CO}_2$  in the integrated samples contained in Tedlar bags were measured on an automated analysis system. Carbon dioxide concentration was measured by a non-dispersive infra-red gas analyser (NDIR, Licor 6251, Licor, Nebraska, USA); methane concentration was measured by a gas chromatograph with a flame ionisation detector (Dani Model 6500, DANI, Italy) on molecular sieve 5A with nitrogen carrier gas; nitrous oxide was measured with a gas chromatograph equipped with an electron capture detector (Hewlett Packard model 5890, Palo Alto, USA) on Porapak QS with 5% methane in argon carrier gas. Both gas chromatographs (GCs) were operated with backflush. Compounds with retention times greater than that of required species were retained on the pre-column and backflushed to the atmosphere in order to minimise the time required and improve the quality of each analysis. Water vapour was removed from all samples by a semi-permeable membrane drier (Nafion, Dupont, USA) prior to analysis. Methane and

nitrous oxide concentrations of the air samples were determined by comparison with a clean air working standard prepared by CSIRO GASLAB using standard protocols linked to the international scales for CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub> (Francey et al., 1993). Multipoint calibrations were performed occasionally to quantify the detector responses over the range of gas concentrations encountered during the experiment. The flame ionisation detector was linear for methane concentration. While the electron conductivity detector was non-linear for nitrous oxide concentration. The analysis system was controlled and logged by a microcomputer. Analytical precision was 0.2% for N<sub>2</sub>O, 0.6% for CH<sub>4</sub>, and 0.2% for CO<sub>2</sub>.

In operation, a flux measurement cycle was initiated by closing the chamber lid. In the integrating mode, gas samples (about 1 litre volume) were withdrawn from the chamber at specified times after closure (commonly 5, 15, 25 and 45 minutes) and accumulated in Tedlar gas sampling bags. One bag was allocated to each of the sampling times. The lid was then opened 2 minutes after the final sample was collected and the chamber was rapidly flushed with ambient air. The lid remained open until the next flux measurement was initiated, when another set of samples were added to the accumulating bags. In the case where a 24-hour average was required, chamber closures were triggered every four hours, and therefore at the end of a 24-hour measurement period each of the four sample bags contained approximately six litres of air. In this study the rate of change in average gas concentration was estimated by either the difference in concentration over one time interval, or by least squares regression of concentration against time where four separate sampling times were used.

### 2.2.2 *Continuous mode – FTIR spectrometry*

Infrared spectra were measured at 1 cm<sup>-1</sup> resolution using a Bomem MB104 FTIR spectrometer fitted with a globar source, InSb detector and 22-m path, 8-L volume White cell (IR Analysis, Anaheim Ca.) housed in a thermostatted enclosure in a mobile laboratory 20 m from the chamber sites. Cell pressure and temperature were monitored and logged along with each spectrum measured. Air from the chamber was circulated continuously through the cell, and spectra were collected continuously and accumulated in 1-minute (20 scans) groups. The entire operation was fully automated under the control of a single Array Basic program running in the GRAMS (Galactic Industries, New Hampshire) spectrometer operating environment. The spectrometer details and methods are similar those described in detail by Esler et al. (2000).

FTIR spectra were analysed by Classical Least Squares (CLS) using calculated calibration spectra as described in detail by Griffith (1996). Measured concentrations could be corrected for cell temperature, pressure and water vapour concentration also retrieved from the FTIR spectra to provide dry air mixing ratios when required. This analysis provided a working calibration scale was related to the GASLAB scale by daily measurements of the clean air working standard used in the GC analyses described above. Analytical precision was better than 0.5ppbv for N<sub>2</sub>O and 1 ppmv for CO<sub>2</sub> for a 1-minute average based on replicate measurements of a single air sample. The analysis showed a small cross-sensitivity of measured N<sub>2</sub>O to CO<sub>2</sub> concentration, of the order of 1 ppbv N<sub>2</sub>O per 100 ppmv CO<sub>2</sub>, due to spectroscopic overlap not fully accounted for by the CLS analysis. This was quantified by measurements where CO<sub>2</sub> was added to the chamber-cell volume at constant N<sub>2</sub>O concentration and used to correct all N<sub>2</sub>O measurements. There is also a similar cross sensitivity to water vapour, but this was less significant because water vapour was

close to saturation at all times and normally did not vary significantly throughout a 30 minute measurement cycle.

Each flux measurement cycle commenced with five minutes of air measurement with the chamber lid open to establish the ambient trace gas concentration. The chamber lid was then closed, and the trace gas concentrations were recorded at 1-minute intervals for the next 20 minutes. The sequence was completed by reopening the chamber for five minutes to flush the chamber with ambient air before the next cycle. In the OASIS experiment the FTIR was switched between two chambers on alternate half hours using a pair of solenoid gas valves under the control of the program (Fig. 2). Thus a flux measurement was made on each chamber each hour, 24 hours per day except for a 0.5-1 hour maintenance break for calibration and data backup.

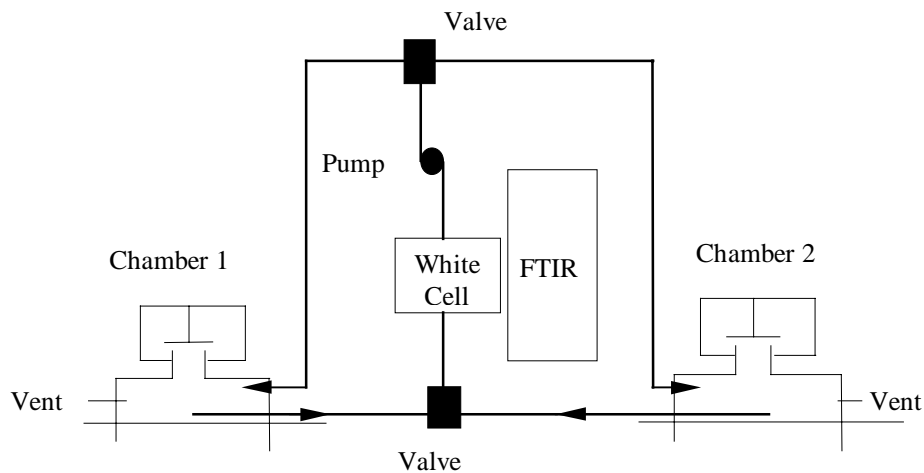


Figure 2 Schematic diagram of the FTIR gas circuit

### 2.3 Site description

The continuous and the integrating methods were used to measure fluxes of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O in two agricultural fields in October 1994 and October 1995 as part of the OASIS campaigns. In both 1994 and 1995, one set of measurements was made in a 15 ha lucerne pasture at the Charles Sturt University farm, near Wagga Wagga, New South Wales (35°4'S, 147°21'E, altitude 219 m). Approximately 70% of the surrounding district is assigned to pasture for grazing sheep and cattle, and the rest is used for crops. Soil at the site grades from a fine sandy loam at the surface (23 g kg<sup>-1</sup> clay) to a mild clay (55 g kg<sup>-1</sup> clay) at 0.6 m with a band of fine iron-manganese nodules (Poss et al., 1995). The second set of measurements was made in an adjacent 15-ha instrumented field to the east planted with oats in 1994 and *Triticale* in 1995.

Six chambers were deployed during each campaign, three in each crop. In order to reduce disturbance to crop growth during the time course of the campaigns, each chamber was supplied with two adjacent bases. The chamber box units were moved to their alternate base each morning ensuring that the measurement sites were covered for a maximum of 24 hours before returning to ambient conditions.

The first season, October 1994, was very dry supporting limited crop growth. Only 154 mm of rain fell in the six months from the beginning of April until field



measurements commenced in October, well below the climatic average of 383 mm. Leaf area indices (LAI) were  $< 0.5$  for the pasture and  $\approx 1$  for the crop in 1994.

For the second season, October 1995, rainfall was above average. Rainfall in the preceding six months was 434 mm, while from 11 to 29 October, LAI increased from 0.6 to 1.5 for the pasture and from 3.1 to 3.8 for the crop.

Significant rainfall events occurred during both the campaigns. In 1994, rainfall occurred on the 27<sup>th</sup> October, at the end of the campaign. The 1995 campaign encompassed a drying cycle spanned by rainfall events on 13<sup>th</sup> and 22<sup>nd</sup> October.

In 1994, only CH<sub>4</sub> and CO<sub>2</sub> exchange was measured. In 1995, N<sub>2</sub>O was included in the measurement suite. Additionally, in 1995 the chamber windows were covered with aluminium foil, and therefore the CO<sub>2</sub> exchange observed during the day was entirely due to soil and plant respiration.

### 3. The chamber trace gas exchange measurement technique

#### 3.1 Principles and assumptions

##### 3.1.1 Chamber mass balance

Fluxes of trace gases are often measured by the closed chamber technique (e.g. Galbally and Roy, 1980; Galbally et al., 1985; Businger, 1986; Kirstine et al., 1998). The principles of the technique, and the artifacts to which it is sensitive, have been reviewed by a number of authors (Denmead, 1979; Hutchinson and Mosier, 1981; Jury et al, 1981; Galbally et al., 1985; Livingston and Hutchinson, 1995; Healy et al., 1996). In brief, the flux of a gas is calculated from the initial rate of change in concentration of the gases of interest when the chamber volume is sealed from the ambient air.

Conservation of mass is applied to the chamber:

$$F = \frac{V}{A} \cdot \frac{dC_t}{dt} + \frac{v_g}{A} (C_t - C_a) + v_d C_t \quad (1)$$

where:

- $F$  is the flux of the trace gas ( $\text{g m}^{-2}\text{s}^{-1}$ )
- $C_t$  is the concentration of the trace gas in the chamber at time  $t$  ( $\text{g m}^{-3}$ ),
- $C_a$  is the concentration of the trace gas in ambient air ( $\text{g m}^{-3}$ ),
- $t$  is the time after closure of the chamber (s),
- $V$  is the volume of the closed chamber ( $\text{m}^3$ ),
- $A$  is the area of ground surface area enclosed by the chamber,
- $v_g$  is the volumetric rate of loss of air from the chamber ( $\text{m}^3 \text{s}^{-1}$ ), described here as the gross rate of loss of air from the chamber,
- $v_d$  is the deposition rate of the gas on the chamber, plant and soil surfaces, ( $\text{m s}^{-1}$ ).

The subscripts  $i$  and  $f$  represent initial and final states respectively.

The gross rate of loss of air from the chamber can be represented by its constituent parts as:

$$v_g = v_s + v_e \quad (2)$$

- $v_s$  is the rate at which air is deliberately withdrawn from the chamber usually for sampling,  
 $v_e$  is the natural ventilation or leakage rate of air from the chamber.

A key feature of this analysis is that the deliberate removal of air and natural leakage of air from the chamber is small such that it makes only a minor contribution to Eq. 1. Choice of a small value of  $t_f$  helps this by minimising  $C_t$ . Sealing the chamber well to minimise natural ventilation is also important.

### 3.1.2 Characterisation of natural air leakage from chambers

The natural ventilation (leakage) rate of the chambers was determined in tests using CO<sub>2</sub> as an inert tracer gas. In these tests, the chambers were located over a sealed anodised aluminium base. A pulse of CO<sub>2</sub> was injected into the chamber immediately after closure of the chamber lid, and the ventilation rate was calculated from the rate of change in the CO<sub>2</sub> concentration in the chamber.

From an analysis of the chamber mass balance, the relationship between the natural ventilation and the time course of tracer concentration can be derived (Marynick and Marynick, 1975; Kimball, 1990), viz.

$$\frac{v_e}{V} = \frac{\ln(C_i / C_f)}{(t_f - t_i)} - \frac{v_s}{V} \quad (3)$$

Measurements of  $v_e/V$  were conducted with all chambers. The results for chamber 5 are presented in Fig. 3 and demonstrate that ambient windspeed has a substantial influence on  $v_e/V$ . It can be demonstrated by applying a mass balance analysis to the chamber via Eq. 1 (Galbally et al., 1985) that a leakage rate of  $v_g/V = 0.1 \text{ h}^{-1}$ , when neglected, leads to an error in the flux of 3.5% for a 45-minute chamber closure. In this study, all fluxes have been corrected for leakage assuming an average chamber leakage rate of  $0.2 \text{ h}^{-1}$ .

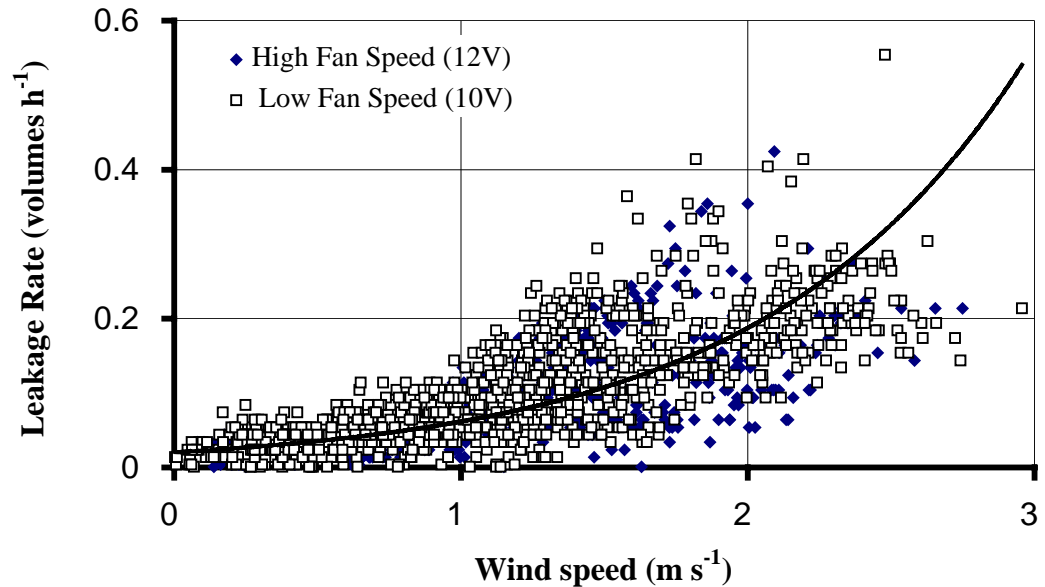


Figure 3 The effect of ambient wind speed on the chamber's leakage rate.

The boundary-layer resistance of these chambers was calculated from measurements of the rate of water evaporation from an underlying fully wet surface into dry air in the chamber, and averaged  $88 \text{ s m}^{-1}$ . It can be shown (Thom and Oliver, 1977) that an aerodynamic resistance of this magnitude is equivalent to a windspeed of  $3 \text{ m s}^{-1}$  at 2 m above a uniform free water surface.

The chambers were also tested for artefact fluxes. The exchanges of trace gases observed over a solid aluminium base, typically  $0.02 \pm 0.03 \text{ ng N m}^{-2} \text{ s}^{-1} \text{ N}_2\text{O}$ ,  $-0.22 \pm 0.20 \text{ ng C m}^{-2} \text{ s}^{-1} \text{ CH}_4$  and  $0.8 \pm 0.3 \text{ } \mu\text{g m}^{-2} \text{ s}^{-1} \text{ CO}_2$ , are negligible compared with the real fluxes observed when there is an open soil/plant surface beneath the chamber. An emission from the soil to the atmosphere is defined as a positive flux.

### 3.1.3 The effect of a deep canopy in the chamber on the concentration time course

Commonly, it is assumed in chamber studies that the air space is well mixed. This assumption will not apply to the case of measurements over a deep dense crop. To our knowledge this situation has not been studied. Consequently, we have undertaken a study using a numerical model of the mixing in the chamber, described in the Appendix, and the results of the modelling are presented here.

Using the theory of Raupach (1987) the variation of both the turbulent diffusion coefficient  $D$  and aerodynamic resistance  $r_a$  with height was calculated for a dense canopy of 1-m depth with similar characteristics to the *Triticale* crop (Fig. 4). This demonstrates that the turbulent diffusion coefficient decreases rapidly towards the bottom of the canopy and approaches the value of the molecular diffusion coefficient of  $\text{N}_2\text{O}$  in ambient air ( $=1.5 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$ ) at the bottom of the canopy. About half of the integrated aerodynamic resistance between the soil surface and the air above the canopy is in the 0.02-m layer above the soil. Therefore  $\text{N}_2\text{O}$  emitted from soil is likely to build up to a much higher concentration at the bottom of the canopy than in the other part.

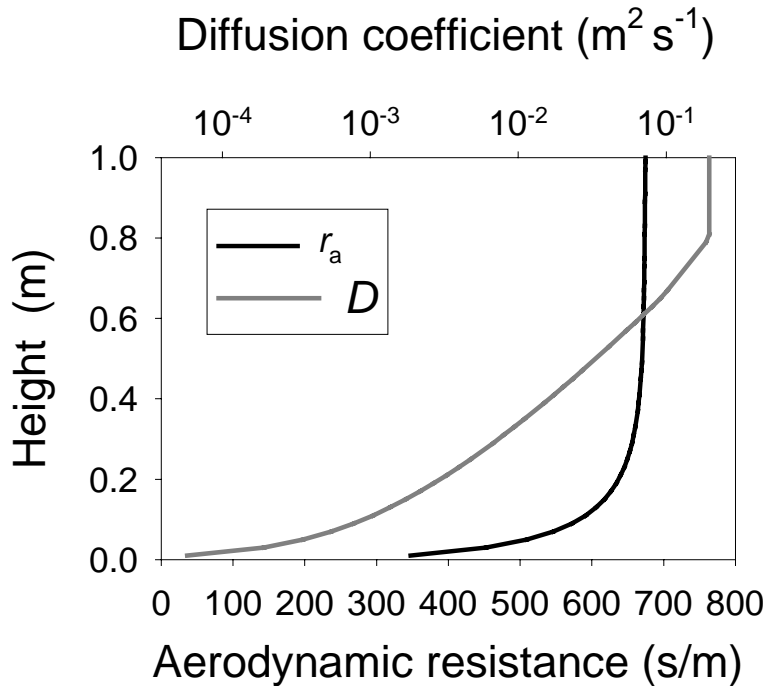


Figure 4 Variation of the turbulent diffusion coefficient and the total canopy resistance from the surface as a function of canopy height for a dense crop of 0.8 m height and leaf area index of 4.

Fig. 5 shows the  $N_2O$  concentration at three different heights within the canopy after the chamber is closed assuming a constant flux of  $10 \text{ ng N m}^{-2} \text{ s}^{-1}$  from the soil at the lower boundary of the canopy. A non-linear increase in  $N_2O$  concentration is most evident at the bottom layer during the first 30 seconds after the chamber closure, because of the poor turbulent mixing. About 60 seconds after the chamber closure,  $N_2O$  concentration within the canopy increases linearly with time and at an equal rate at all heights, therefore flux calculated from air samples taken at 600 and 1800 seconds after chamber closure will provide an unbiased estimate of the soil  $N_2O$  emission. This result is also valid if the soil  $N_2O$  flux at the bottom of the canopy varies with time, as long as it is not directly dependent on  $N_2O$  concentration at the surface through metabolic feedback mechanisms. Therefore trace gas transport through the canopy is unlikely to introduce errors into the flux estimates despite the occurrence of a significant concentration gradient. Thus complete mixing within the air space of the chamber is not always a necessary condition for valid chamber measurements.

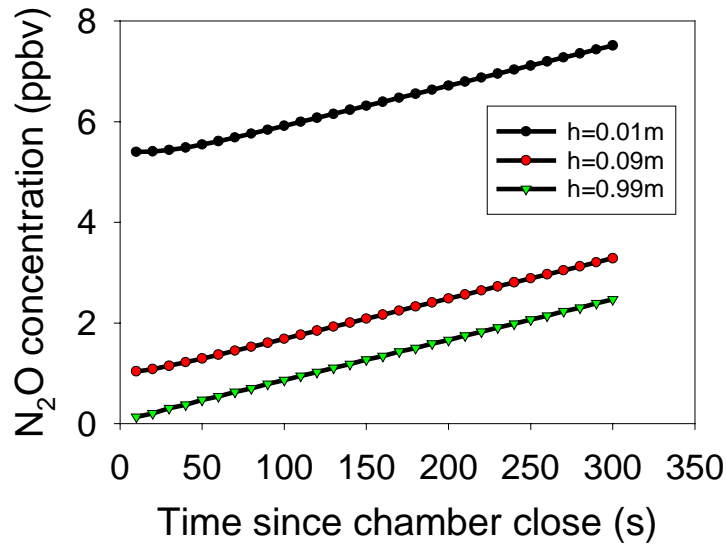


Figure 5 Simulated time course of trace gas concentration near the soil surface, at 0.01m, 0.09 m and at 0.99 m above the soil surface for an N<sub>2</sub>O flux of 10 ng N m<sup>-2</sup> s<sup>-1</sup> into a static chamber for a dense crop of 1m and leaf area index 4. In the real world these concentrations would represent the perturbations of nitrous oxide concentration above the prevailing ambient concentration.

The details presented so far show that the flux chamber method has a rigorous basis and is constructed so that the soil/plant/atmosphere exchange processes in the chamber mimic those in the outside world as closely as possible. The system has operated successfully in the field on many occasions providing extended periods of continuous trace gas exchange measurements for CO<sub>2</sub> from respiration, N<sub>2</sub>O and CH<sub>4</sub> (Meyer et al., 1997, Wang et al., 1997).

### 3.2 Integrated and continuous flux measurement methods

When there is no loss of the gas to the chamber itself and no back reaction with the source on earth, then the rate of change in trace gas concentration is:

$$F = \frac{V}{A} \frac{dC}{dt} + \frac{v_g}{A} (C_t - C_a). \quad (4)$$

Equation 4, in time transformed units, is used for the derivation of fluxes. This equation can only be fitted to the data using non-linear techniques (Galbally et al., 1985). An alternative to this is a solution to Eq. 4 that corrects for chamber leakage (e.g. Marynick and Marynick, 1975), by modifying time  $t$  using the transform

$$t' = \frac{(1 - e^{-v_g V^{-1} t})}{v_g V^{-1}} \quad (5)$$

where  $t'$  is the leak-corrected time. This transform collapses the second term from the right-hand side of Eq. 4 into the first term on the right-hand side. The formulation used involves the linear regression of  $C$  versus  $t'$ . Under these circumstances Eq. 4 can be reduced to a simple derivative or finite difference form:

$$F = \frac{V}{A} \cdot \frac{dC}{dt'} = \frac{V}{A} \cdot \left( \frac{C_f - C_i}{\Delta t'} \right) \quad (6)$$

where the subscript  $\Delta t'$  is the time interval, ( $\Delta t = t_f - t_i$ ) expressed in transformed units. The values of  $t_i$  and  $t_f$  are commonly 5 and 45 minutes. This approach is used for the continuous method.

The following provides the basis of the integrating method. The mean daily flux rate  $\bar{F}$ , calculated from  $n$  individual flux measurements,  $F_j$  is

$$\bar{F} = \left[ \sum_{j=1}^{j=n} F_j \right] / n. \quad (7)$$

If the sampling times for each individual measurement ( $t_i, t_f$ ) are constant throughout the experiment (as is the case with automatically controlled chambers), and a constant air mass is withdrawn from the chamber at each sampling (as is approximately the case with voltage regulated pumps and small ambient temperature variation), then substituting Eq. 6 into Eq. 7 gives

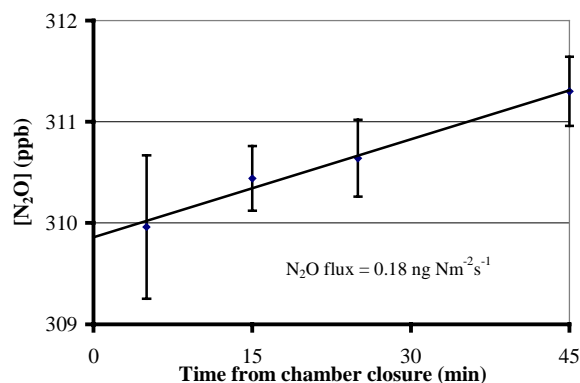
$$\bar{F} = \frac{V}{A\Delta t} \left( \frac{\sum_{j=1}^{j=n} C_{fj}}{n} - \frac{\sum_{j=1}^{j=n} C_{ij}}{n} \right) \quad (8)$$

or

$$\bar{F} = \frac{V}{A\Delta t'} (\bar{C}_f - \bar{C}_i) \quad (9)$$

where  $\bar{C}_f$  and  $\bar{C}_i$  are, respectively, the average concentrations of the  $n$  initial and  $n$  final air samples. In turn,  $\bar{C}_f$  and  $\bar{C}_i$  can each be calculated either from  $n$  individual chemical analyses or, if the  $n$  samples are of equal mass and are mixed prior to analysis, from a single analysis of the bulked air sample. Thus the average flux obtained from the set of individual flux values can be obtained from the measurement of the average concentrations as represented by the accumulated initial and final air samples. This is the basis of the integrated chamber technique. There were no 24-h (4 bag) timecourses recorded using the integrating method over equilibrated soil sites in the OASIS study; however a typical example of a timecourse measured using the integrated method in related study conducted in a Eucalyptus forest in southern Victoria (Meyer et al.; 1997) is shown in Fig. 6.

A.



B.

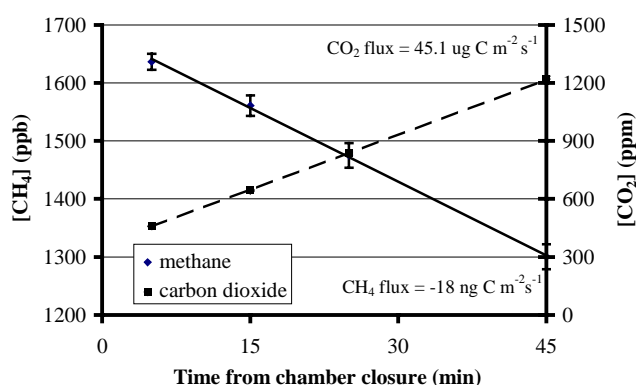
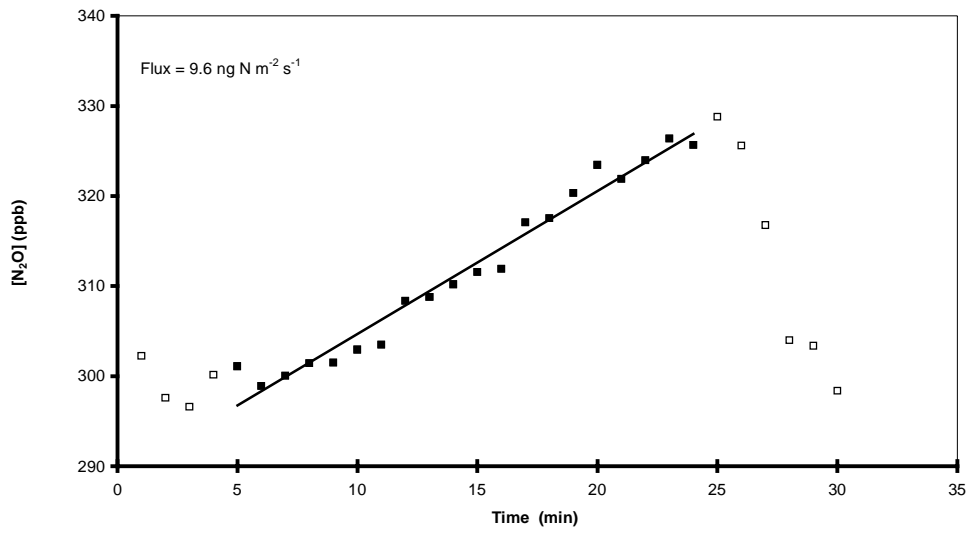


Figure 6. Time-course of (a) N<sub>2</sub>O concentration and (b) CH<sub>4</sub> concentration (diamonds) and CO<sub>2</sub> concentration (squares) in the chamber headspace during the 24-h mean soil-atmosphere flux measurement in a Eucalyptus forest near Spargo Creek, Victoria. The data points are mean  $\pm$  standard errors of the mean for five replicate analyses of the bulked air samples. In the case of CO<sub>2</sub> the standard error bars are smaller than the data points. The lines are the least-squares linear regressions.

Typical examples of N<sub>2</sub>O and CO<sub>2</sub> concentration time courses measured by FTIR using the continuous method are shown in (Fig. 7). In all cases, the time courses were linear. Though in principle methane fluxes can also be determined by FTIR, in practice the variability in the methane concentration observed by FTIR was too large for the calculation of meaningful methane fluxes. Measurements with the FTIR when the chamber remained closed for 7 hours (transformed time) showed no non linearity in the rates of change of N<sub>2</sub>O and CO<sub>2</sub> concentration, see (Fig. 8).

A.



B.

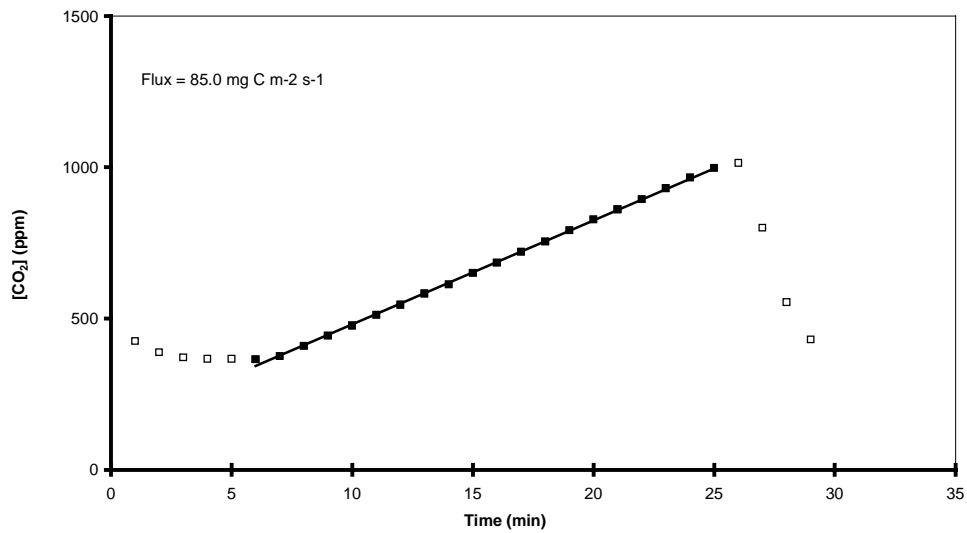


Figure 7 Time course of trace gas concentration and derived emission rates on 24 October 1995 at 1430 local time measured by FTIR. Chamber open, open symbols; chamber closed, closed symbols. (a)  $\text{N}_2\text{O}$  (b)  $\text{CO}_2$



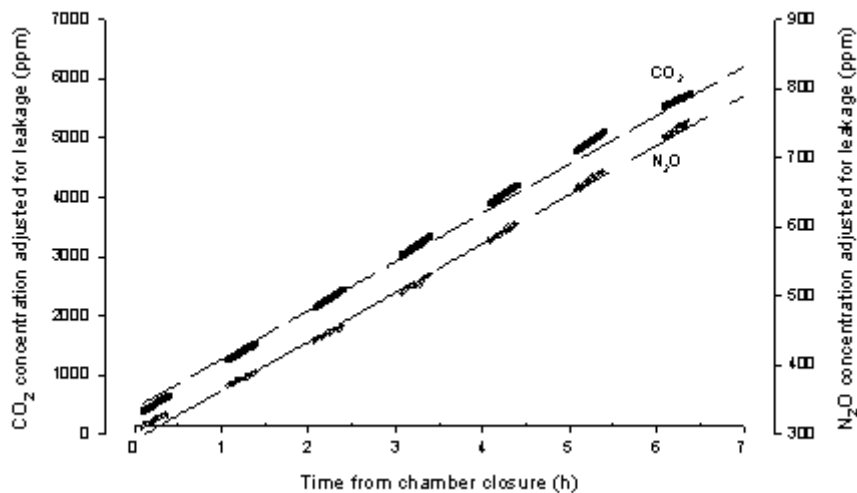


Figure 8 Timecourse of CO<sub>2</sub> and N<sub>2</sub>O concentration during a chamber flux measurement over lucerne pasture. The flux chamber remained closed for seven hours. Times are corrected for a chamber air exchange rate of 0.2 h<sup>-1</sup>.

## 4. Field observations

### 4.1 Time series of CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub>

#### 4.1.1 Integrating method

The average daytime, night-time and daily fluxes of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O observed during the 1994 and 1995 campaigns are summarised in Table 1. Respiratory carbon dioxide fluxes observed in 1995 were more than double those observed in 1994 and showed a strong diurnal variation with significant differences between the pasture and *Triticale* crop, Figs. 9a and 10a. Night-time fluxes averaged 35 and 44  $\mu\text{g C m}^{-2} \text{s}^{-1}$  in the lucerne and *Triticale* paddocks respectively while daytime soil plus plant respiration averaged 58 and 94  $\mu\text{g C m}^{-2} \text{s}^{-1}$  in the lucerne and *Triticale* respectively. The night-time and day-time fluxes of CO<sub>2</sub> and N<sub>2</sub>O observed during 1995 using the integrating method are shown in Fig. 10. The difference between day-time and night-time fluxes increased between the two rain days, 13<sup>th</sup> and 22<sup>nd</sup> October, in concert with air temperature. This was particularly apparent for the *Triticale* crop where the daytime respiratory CO<sub>2</sub> flux exceeded the night-time flux by a factor of 1.2 to 3.5 in contrast to the lucerne pasture where the diurnal variation ranged from a factor of 1.1 to 2. There was no indication of any stimulation in respiration immediately following rain, suggesting that soil and plant respiration were not water limited. This contrasted with 1994 when a minor rainfall event stimulated a 3 to 4-fold increase on CO<sub>2</sub> emission from both oats and pasture (data not presented). Site-to-site variability for CO<sub>2</sub> exchange was small, which implies that the averaged fluxes should give reasonably good comparative estimates for each crop.

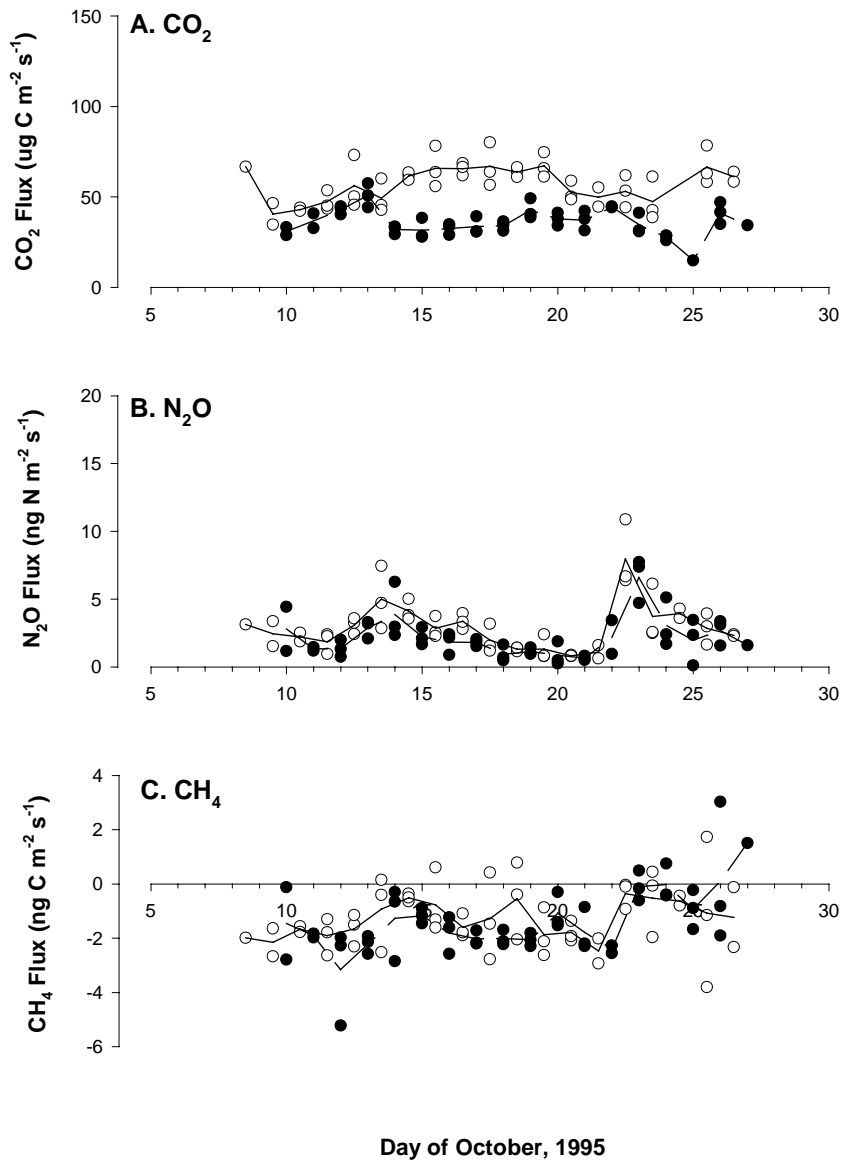


Figure 9 Time course of (a) CO<sub>2</sub> soil plus plant respiration, (b) N<sub>2</sub>O and (c) CH<sub>4</sub> soil emission measurements from the lucerne pasture using the integrating method. Individual chamber measurements are shown as points and averages for all chambers are shown as lines for daytime (open symbols, solid line) and night-time (closed symbols, dashed line) fluxes.

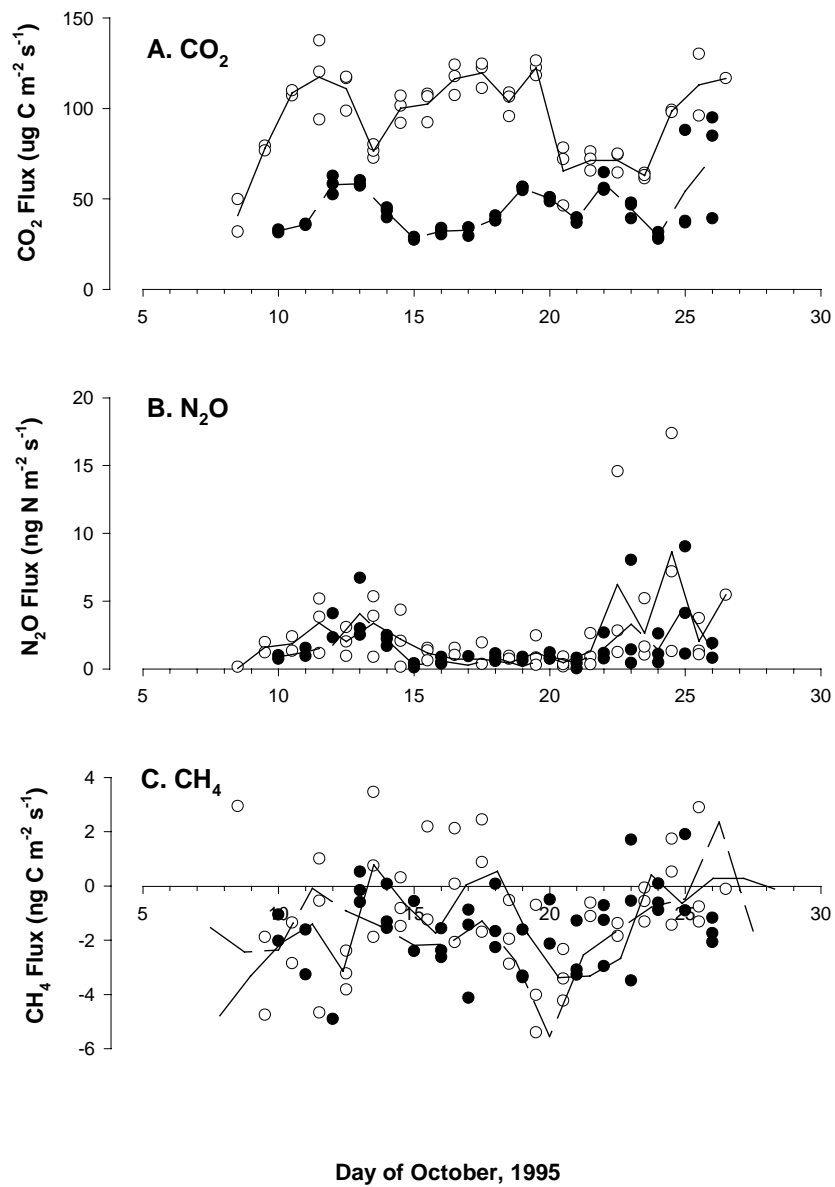


Figure 10 Time course of (a) CO<sub>2</sub> soil plus plant respiration, (b) N<sub>2</sub>O and (c) CH<sub>4</sub> soil emission measurements from the Triticale crop using the integrating method. Individual chamber measurements are shown as points and averages for all chambers are shown as lines for daytime (open symbols, solid line) and night-time (closed symbols, dashed line) fluxes.

Table 1 Average daily fluxes of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O from all soil sites measured during the OASIS campaigns in October 1994 and October 1995. The CO<sub>2</sub> fluxes constitute soil plus plant respiration only. Means ± standard errors of the mean. By definition, a positive flux is an emission from the soil to the atmosphere.

Crop		Day-time	Night-time	24-hour
<b>1994</b>				
<i>Lucerne</i>	CO <sub>2</sub> flux (µg C m <sup>-2</sup> s <sup>-1</sup> )	-	17 ± 4	-
	CH <sub>4</sub> flux (ng C m <sup>-2</sup> s <sup>-1</sup> )	-4.9 ± 1.1	-4.1 ± 0.5	-5.5 ± 0.9
<i>Oats</i>	CO <sub>2</sub> flux (µg C m <sup>-2</sup> s <sup>-1</sup> )	-	29 ± 6	-
	CH <sub>4</sub> flux (ng C m <sup>-2</sup> s <sup>-1</sup> )	-3.9 ± 0.8	-5.9 ± 0.5	-5.3 ± 0.4
<b>1995</b>				
<i>Lucerne</i>	CO <sub>2</sub> flux (µg C m <sup>-2</sup> s <sup>-1</sup> )	60 ± 2	38 ± 1	48 ± 3
	CH <sub>4</sub> flux (ng C m <sup>-2</sup> s <sup>-1</sup> )	-1.4 ± 0.2	-1.4 ± 0.3	-1.5 ± 0.2
	N <sub>2</sub> O flux (ng N m <sup>-2</sup> s <sup>-1</sup> )	3.1 ± 0.4	2.3 ± 0.4	2.9 ± 0.4
<i>Triticale</i>	CO <sub>2</sub> flux (µg C m <sup>-2</sup> s <sup>-1</sup> )	100 ± 6	48 ± 3	77 ± 4
	CH <sub>4</sub> flux (ng C m <sup>-2</sup> s <sup>-1</sup> )	-1.5 ± 0.4	-1.7 ± 0.4	-1.5 ± 0.3
	N <sub>2</sub> O flux (ng N m <sup>-2</sup> s <sup>-1</sup> )	2.5 ± 0.5	1.6 ± 0.4	2.1 ± 0.4

The nitrous oxide fluxes exhibited considerable day-to-day variation, with maximum emission rates occurring immediately after rainfall, followed by a slow decline during the subsequent 10 days as the soil dried, Fig. 9b and Fig. 10b. The maximum 12-h N<sub>2</sub>O flux observed from the lucerne pasture was 11 ng N m<sup>-2</sup>s<sup>-1</sup> following rain on 22<sup>nd</sup> October rising from a minimum of 1 ng N m<sup>-2</sup>s<sup>-1</sup> on 21<sup>st</sup> October. The N<sub>2</sub>O fluxes from the *Triticale* crop on average were approximately 25% lower than the fluxes from the lucerne pasture and followed a similar time course. The maximum N<sub>2</sub>O flux observed in the *Triticale* was 18.1 ng N m<sup>-2</sup>s<sup>-1</sup> on 24<sup>th</sup> October. The *Triticale* sites showed far greater variability in response of N<sub>2</sub>O flux to the second rainfall event than was apparent in the lucerne with little response evident in chamber 4, a similar response to the lucerne shown in the chamber 5 sites, and a delayed response observed in the chamber 6 sites. This variability, which was not mirrored in either the CH<sub>4</sub> or CO<sub>2</sub> fluxes, indicates greater heterogeneity in the distribution of anaerobic soil sites in the *triticale* crop than in the lucerne crop, possibly a consequence of the greater water demand by the *Triticale* crop. Daytime N<sub>2</sub>O fluxes were slightly larger than the night-time fluxes, particularly on the fine days following the rain on the 13<sup>th</sup> October.

Methane fluxes in 1995 were 75% lower than those recorded from the same sites during the drought year of 1994, Table 1, averaging approximately 1.5 ng C m<sup>-2</sup>s<sup>-1</sup> with little difference between daytime and night-time fluxes. Further there was considerable day-to-day variation in methane oxidation rate particularly in the second wetter year, 1995. In both the *Triticale* and lucerne paddocks, methane uptake rate was smallest immediately following rain and largest at the end of the drying cycle.

#### 4.1.2 Continuous measurements.

The time course of hourly measurements of carbon dioxide exchange for a single chamber in the lucerne pasture, (Fig. 11), reveals a daily cycle of large amplitude that was highly correlated with temperature, as shown in (Fig. 12). A simple exponential relationship explains most of the variability ( $r^2 = 0.83$  for lucerne and 0.92 for *Triticale*), which supports the conclusion drawn earlier from the integrated flux measurements that respiration was controlled principally by temperature and was not limited by soil water.

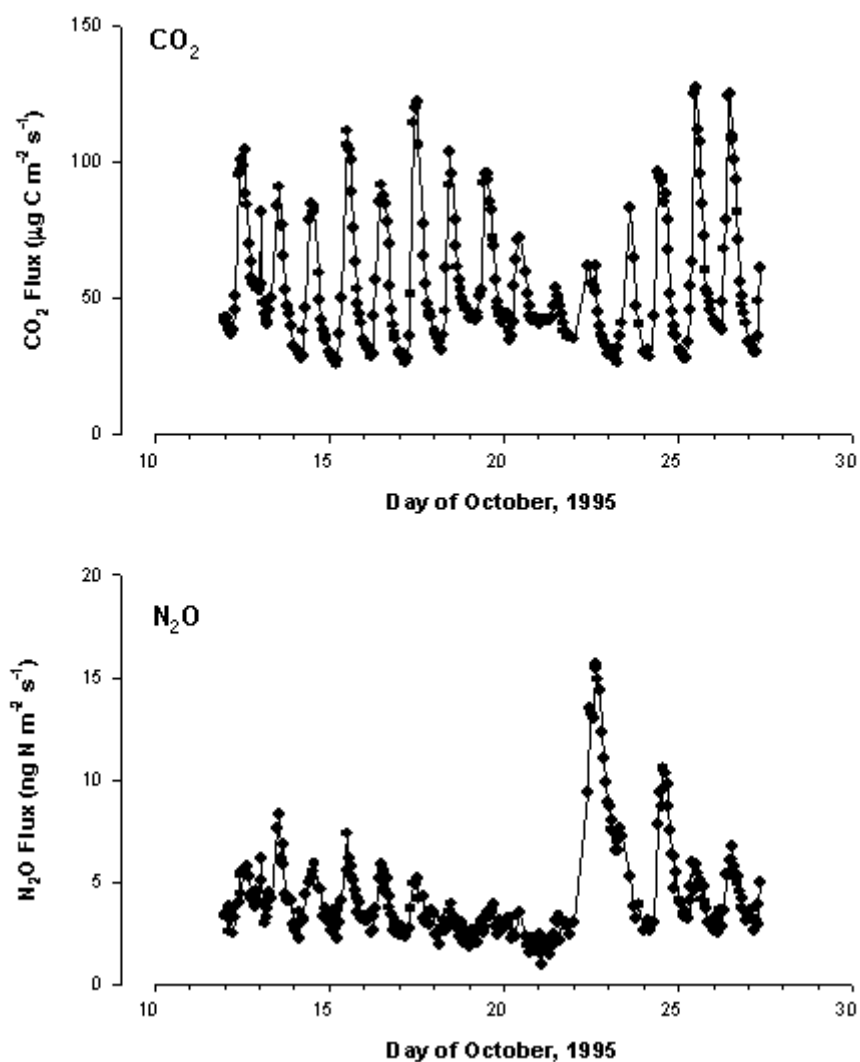


Figure 11 Time course of (a) CO<sub>2</sub> soil plus plant respiration and (b) N<sub>2</sub>O soil emission measurements from the lucerne pasture using the continuous mode chamber method coupled to FTIR gas analysis.

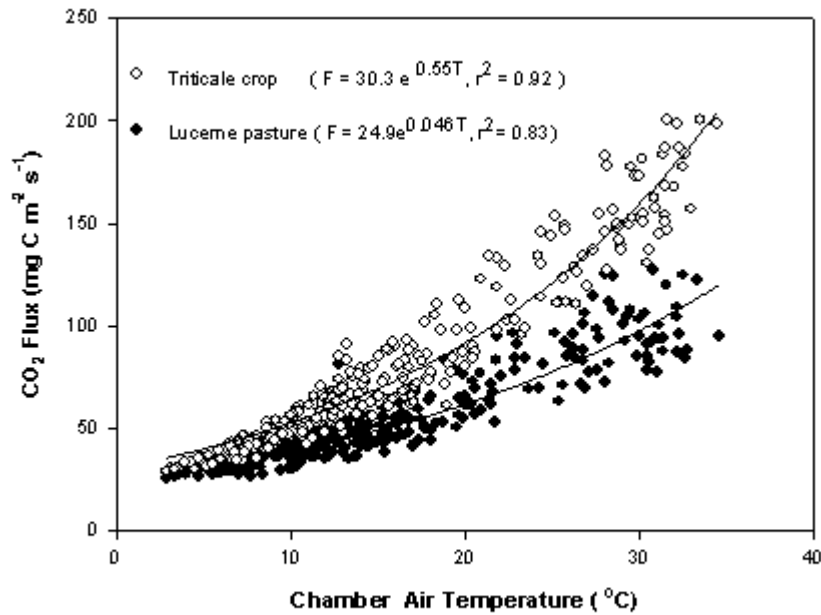


Figure 12 Relationship between soil plus plant dark respiration and air temperature. Hourly fluxes, measured by FTIR. Open symbols, *Triticale* crop; closed symbols, lucerne pasture both for October 1995.

The time course of hourly nitrous oxide flux from the lucerne pasture site is also shown in (Fig. 11). Both rainfall events on October 13 and 22 clearly stimulated a pulse of N<sub>2</sub>O emission and increased the sensitivity of this N<sub>2</sub>O emission to diurnal temperature variation. However, these effects were transient, and within 7 days N<sub>2</sub>O emission had declined to a constant basal rate. Large but transient responses to rainfall are well known properties of denitrification events and suggest that the short-term variability in N<sub>2</sub>O emission is due mainly to denitrification.

#### 4.1.3 Discussion of time series

The salient features of these chamber measurements may be summarised as follows. When both crop and pasture paddocks had adequate soil moisture, respiratory CO<sub>2</sub> fluxes were significant, and controlled largely by temperature. Nitrous oxide fluxes were dependent on both rainfall events and temperature, though the temperature sensitivity appeared to be moisture dependent. Methane oxidation rate responded inversely to soil moisture content, with the rate of change greatest as the soil moisture level approached field capacity.

The fluxes observed at Wagga are broadly consistent with observations from other agricultural systems in Australia, and comparable nitrogen limited systems in the USA. The night-time CO<sub>2</sub> fluxes were similar to those observed in natural temperate forest soils in southern Australia and grazed pasture near Wagga Wagga (Galbally et al., 1995; Wang et al., 1997). The nitrous oxide fluxes observed in this experiment were also consistent with the rates of 1-2 ng N m<sup>-2</sup> s<sup>-1</sup> reported from several agricultural sites in south eastern Australia and tended toward the lower end of the emission rates reported from regularly fertilised cropland in Europe and USA (Bouwman, 1990).

Recently, there has been considerable interest in the effect on methane exchange of soil disturbance by agricultural activities. Where direct comparisons can be made between natural and agricultural systems, there is clear evidence that the conversion of undisturbed lands to agricultural activity may suppress the methane uptake (Mosier et al., 1991, 1993, 1997; Ojima et al., 1993; Willison et al., 1995) or even reverse it (Keller and Reiners, 1994; Steudler et al., 1996). Ojima et al. (1993) estimated that in the USA conversion of natural soils to agriculture coupled with nitrogen fertilization has reduced methane uptake by soils by 30 to 75%, and that, globally, agriculture has reduced the methane sink by 30%. A comparison between the methane fluxes found in this study (1.5 to 5.5 ng C m<sup>-2</sup>s<sup>-1</sup>) and those from temperate forests (10 to 16 ng C m<sup>-2</sup>s<sup>-1</sup>) is consistent with this conclusion.

The comparison between the temperature dependence of respiration in the two crops suggests that the *Triticale* crop was more sensitive to air temperature than the lucerne pasture. The relationship between carbon dioxide flux and air temperature yields Q<sub>10</sub> values, (i.e. the ratio of the respiration rates at 10° C intervals) of 1.6 and 1.8 for the lucerne pasture and the *Triticale* crop respectively. These values, though consistent with measurements in other ecosystems (e.g. Rayment and Jarvis, 1997), are lower than the value of 2 that is characteristic of purely metabolic processes. The lower values indicate the presence of some physical constraints.

The cause of the difference in temperature sensitivity between the two crops may be the difference in plant biomass. The above-ground biomass in the *Triticale* crop was considerably larger than the lucerne in the pasture and therefore above-ground respiration contributes a greater proportion of the total CO<sub>2</sub> flux in the *Triticale* than in the pasture. Above-ground plant respiration is strongly coupled to air temperature. Soil and root respiration rates are determined by soil temperature. However, since the amplitude of the daily temperature cycle is increasingly attenuated with soil depth, using air temperature as a surrogate for soil temperature will tend to underestimate the temperature dependence of soil respiration. Hence pasture, in which soil and plant respiration contribute significantly to the total respiratory fluxes, will apparently be less sensitive to air temperature changes than crops in which above-ground plant respiration is more dominant.

Soil porosity is perhaps the most important of the factors affecting nitrous oxide production and methane oxidation rate (Keller and Reiners, 1994; Castro et al., 1995; Mosier et al., 1996; Steudler et al., 1996). Rainfall events that partially saturate soil pore space create the anaerobic conditions required for denitrification. At the same time, methane oxidation rate is reduced by an increase in the resistance to methane diffusion from atmosphere to the oxidation sites in the soil. Both the difference in methane oxidation rate between 1994 and 1995 and the variation in methane flux during the drying cycle of 1995 are consistent with the difference in soil water content.

In general, therefore, the first two requirements for validation have been met, at least qualitatively. The magnitude of the fluxes observed with both techniques are consistent with the fluxes observed independently in comparable agricultural systems, and the short-term responses to environmental drivers are consistent with the known behaviour of soil microbial and physical processes.

## 4.2 Comparison of the continuous method with the integrating method

During the 1995 OASIS campaign both techniques were run alternately using the same two chambers on the same soil/plant plots. The continuous mode flux measurements alternated with the integrated mode measurements, every 30 minutes. The high frequency FTIR trace gas exchange measurements were averaged to produce fluxes for the same integration period as the integrated measurements. The two systems should give identical results if the following requirements are met:

- air mixing in the chambers is adequate;
- the system is inert to trace gases;
- the differential air pressure between the chamber and ambient is identical in each configuration;
- the effect of the chamber on the soil/plant environment is identical;
- the air flows out of the chamber are accurately determined;
- the volumes of both systems are accurately determined;
- both trace gas measurement systems are calibrated against a common standard;
- the responses of the two chemical analytical instruments are identical for the matrix of chamber air that contains the gases being analysed; and
- the time bases for the both sets of measurements are identical.

A detailed examination suggests that all of these requirements were met in the current system configuration. The major difference between the systems was the recirculating pump in the FTIR circuit.

The issue of mixing deserves comment; because there is a physical difference in the characteristics of all mixing in the two chambers. The continuous mode includes an air return of  $15 \text{ l min}^{-1}$ . This emerges from an orifice of 0.1 cm cross-section with a velocity of more than  $20 \text{ m s}^{-1}$ . This air flow will create an additional source of turbulent mixing and stirring within the chamber. The  $\text{CO}_2$  emissions arise from the soil and plants and, in a deep canopy, the above ground plant tissue is probably the major source. In contrast, the  $\text{N}_2\text{O}$  fluxes derive from the soil. Therefore, the characteristics of  $\text{CO}_2$  and  $\text{N}_2\text{O}$  transport through the canopy to the chamber headspace might differ. However, as previously discussed in Section 3.1.3, incomplete mixing in the chamber does not bias the flux measurements.

In practice, though the two methods yielded largely consistent results, there were some differences. The average day and night  $\text{CO}_2$  fluxes measured by the integrating and the continuous methods are compared in Fig. 13a and Table 2. The analysis using a reduced major axis regression, shows no significant differences between the two methods with either the small chamber on the lucerne crop or the large chamber on the *Triticale* crop. The intercepts are less than 10% of the mean fluxes and are not significantly different from zero.



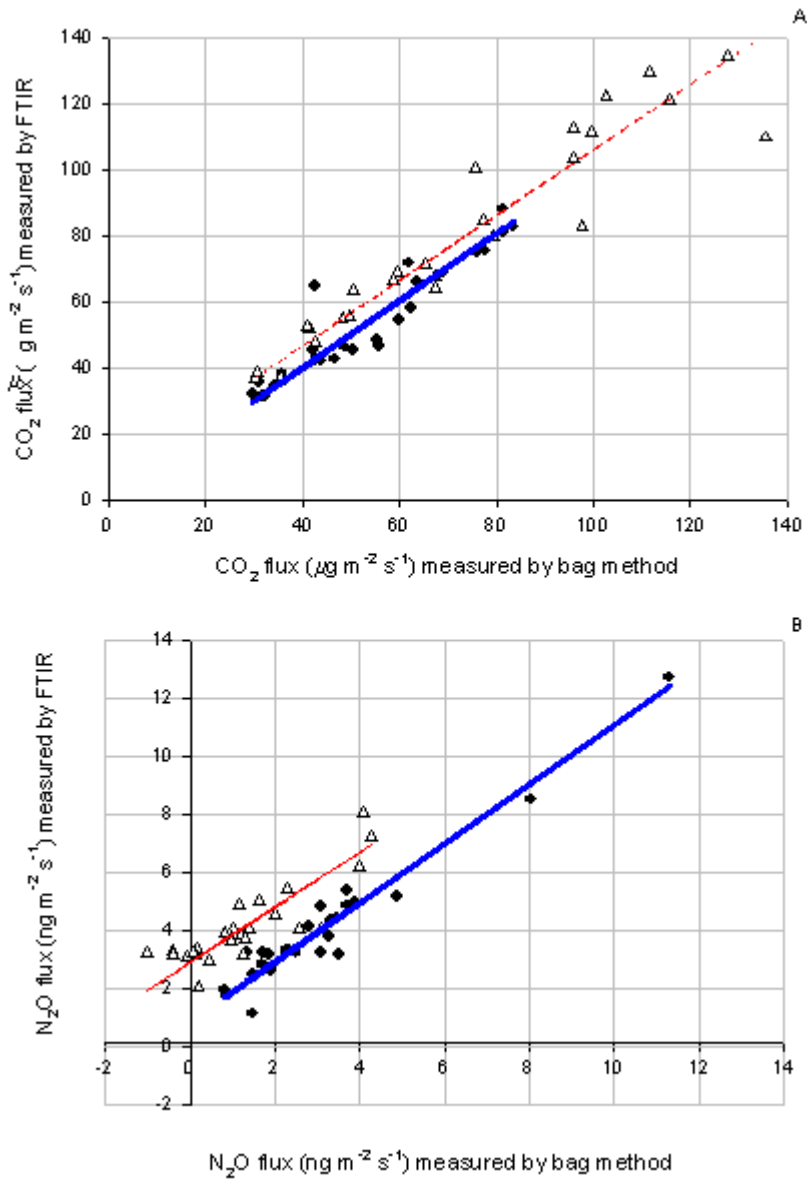


Figure 13 Comparison measurements of (a) CO<sub>2</sub> fluxes and (b) nitrous oxide fluxes by the integrating method and by the continuous method averaged over comparable time intervals. Closed symbols and solid line, lucerne pasture; open symbols and dashed line, *Triticale* crop, both for October 1995.

Table 2 Regression coefficients for a reduced major axis regression between the flux estimates of CO<sub>2</sub> and N<sub>2</sub>O made by the integrating (bag) method and the continuous (FTIR) method averaged over identical time intervals.

Crop	CO <sub>2</sub> flux* ( $\mu\text{g C m}^{-2} \text{s}^{-1}$ )		N <sub>2</sub> O flux* ( $\text{Ng N m}^{-2} \text{s}^{-1}$ )	
	Slope	Intercept	Slope	Intercept
Lucerne Pasture	$1.02 \pm 0.13$	$-0.7 \pm 7.8$	$1.03 \pm 0.0.10$	$0.85 \pm 0.38$
<i>Triticale</i>	$0.99 \pm 0.13$	$7.2 \pm 10.4$	$0.95 \pm 0.20$	$2.9 \pm 0.38$

\* coefficient  $\pm$  95% confidence limits.

The comparison between the N<sub>2</sub>O flux measurements by the two methods Fig. 13b, shows that the integrating method underestimated the flux by 20% with the small chamber on lucerne pasture and 60% with the larger chamber on the *Triticale* crop relative to the FTIR/continuous method, for mid-range fluxes of about  $5 \text{ ng N m}^{-2} \text{ s}^{-1}$ . These differences were due entirely to a significant positive offset between the two methods. This offset was larger with the larger chamber used on the *Triticale* crop i.e.  $2.9 \pm 0.4 \text{ ng N m}^{-2} \text{ s}^{-1}$  compared with  $0.9 \pm 0.3$  for the small chamber on the lucerne crop (Table 3). In both cases the offset is significant relative to the average fluxes observed during the experiment.

There are several plausible explanations for the differences between the two methods. These include:

1. The mixing in the chamber in the dense high crop (*Triticale*) may be inadequate in the case of the integrated system, but better due to the return flow in the continuous mode. (The chamber was not designed to hold such a crop, and the mixing under such conditions is unknown). If the mixing was inadequate in the integrated system the emissions of N<sub>2</sub>O and CO<sub>2</sub> could be suppressed by the build-up of high concentrations of these gases in the bottom of the chambers.
2. The GC and FTIR are subject to cross sensitivity particularly for gases such as CO<sub>2</sub> and N<sub>2</sub>O. Though the FTIR was explicitly calibrated for cross sensitivity between N<sub>2</sub>O and both CO<sub>2</sub> and H<sub>2</sub>O, the issue deserves further investigation.
3. If, despite the vent, there was significant pressure variation in the chamber caused by circulating air through the FTIR gas circuit, then the trace gas fluxes might be increased by the effect of pressure pumping on the mass transport of trace gas from the soil. This phenomenon is distinct from the diffusive and turbulent transport within the canopy discussed in section 3.2. It has been reviewed in detail by Livingston and Hutchinson (1995), has been modelled by several authors (e.g. Massman et al., 1997) and studied both experimentally and theoretically by Gao and Yates (1998a, 1998b), and has been shown to increase apparent flux rates significantly.

The first explanation is unlikely because suppression of trace gas emission from the soil due to an increasing concentration in the atmosphere would lead to a nonlinear time course of the trace gas concentration. The concentration time courses for CO<sub>2</sub> and N<sub>2</sub>O recorded with the continuous method, after correction for a leakage rate of  $0.2 \text{ h}^{-1}$ , were always linear, even on one occasion when, due to a timing error, the chamber remained closed for seven hours. The concentration changes during this event, 2500 ppm CO<sub>2</sub> and 100 ppb N<sub>2</sub>O, were very much greater than the

concentration gradients that might be supported by a dense canopy, see Fig. 5, and yet there was no indication that the flux of either species was suppressed by the increasing concentration in the chamber headspace. Our modelling study, section 3.2, and the Appendix show that on its own, turbulent and diffusive transport through a canopy will not induce non-linearities or biases in the concentration timeseries that might lead to errors in flux determinations.

Cross sensitivity between N<sub>2</sub>O and another unidentified trace gas emitted by the canopy gases could lead to an offset proportional to canopy size. While cross sensitivities can occur in the FTIR analytical technique, known cross sensitivities were entirely accounted for in the analysis of the residuals. There were no other gases that absorbed significantly in the region of the infra red spectrum used for N<sub>2</sub>O and CO<sub>2</sub> detection.

While it is probable that the continuous method induced pressure fluctuations in the chamber by the return flow from the pump, it would be expected that a larger chamber volume would dampen the fluctuations to a greater extent than a smaller chamber volume and therefore the offset in fluxes measured by the two methods would be inversely proportional to chamber volume. However, the offset appeared to increase with increasing chamber volume. On the other hand, the pressure fluctuations might also significantly enhance the diffusion of trace gas through the plant canopy and upper layers of the soil profile. This process was not included in our model in section 3.2, or in the analyses of Healy et al. (1997) and Hutchinson et al. (2000). The deep and dense canopy of the *Triticale* crop would be affected to a greater extent than the smaller lucerne canopy. Further, CO<sub>2</sub> and N<sub>2</sub>O diffusion might be affected differently because a significant proportion of the CO<sub>2</sub> flux derives from leaves in the upper layers of the canopy, in contrast to N<sub>2</sub>O, which is produced exclusively in the soil. Massman et al. (1997) modelled the influence of pressure pumping on trace gas diffusion through a snowpack overlying soil and concluded that the effects of pressure pumping diminished with the magnitude of the concentration gradient, and were essentially transient. Gao and Yates (1998a, 1998b) have shown for continuous flow chambers that the flux can be perturbed to either a lower or higher flux value, in some cases differing by a factor of 5 from the static chamber observed value and the theoretically estimated value. We surmise that some unknown aspect of the flow generated the systematic differences in fluxes observed here in the FTIR observations.

## 5. Conclusions

The integrating method and the continuous-mode static chamber methods have been used during the OASIS experiment to provide a comprehensive estimate of N<sub>2</sub>O, CH<sub>4</sub> and respiratory CO<sub>2</sub> fluxes for comparison with coincident measurements made by micrometeorological methods. The theoretical and experimental basis of these chamber techniques is presented.

Qualitatively, the CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub> flux rates observed during the OASIS experiment by the two independent chamber methods appear to be consistent in magnitude to rates measured in other similar agricultural systems. The variation in fluxes in response to major environmental factors also appears to be consistent with well described soil metabolic processes, and will be analysed quantitatively in a subsequent study.

The comparison between the two flux methods showed that for the most part the results were comparable. That is, the CO<sub>2</sub> emission rates were equivalent and the comparison of the N<sub>2</sub>O emission showed that the two techniques differed by a constant that was independent of N<sub>2</sub>O flux. The cause of the offset was not identified, however several possibilities were investigated and eliminated. These included: restrictions to diffusive transport through a dense canopy, cross sensitivity in the FTIR between N<sub>2</sub>O and an other known trace gases, and time lags caused by the large volume of the FTIR cell. However, one potential artefact could not be eliminated was disturbance to the trace gas concentration profile in the soil by small pressure fluctuations induced by recirculating analysis air. This possibility justifies further investigation.

The systematic offset between the two methods with respect to N<sub>2</sub>O flux is significant because the size of the offset is similar to the size of the average fluxes being measured. We consider that this systematic offset most probably results from an artefact in the continuous method, and therefore the integrating method delivers a better estimate of the absolute fluxes. Nevertheless, direct comparison of the N<sub>2</sub>O fluxes between the crops remains difficult until the source of the offset is unequivocally established. Fortunately both methods deliver the same response to change in N<sub>2</sub>O flux (i.e. the slope of the regression is 1), therefore conclusions about the relative variation of N<sub>2</sub>O exchange within each crop can be made with confidence. Similarly if comparisons are confined to the relative variation in flux rate, then the two methods also deliver complementary spatial and temporal information for N<sub>2</sub>O exchange. Further comparison of flux measuring techniques is desirable.

## **6. Acknowledgements:**

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## Appendix: The effect of a dense plant canopy in the chamber on the concentration time course

As shown in Eq. 1 the closed chamber provides a measurement of the flux from the soil through a measurement of the time rate of change of the volume-averaged concentration of gas under consideration in the chamber.

It is assumed that the canopy aerodynamic resistance was much smaller than the soil resistance and so does not have significant effect on the flux measurements. This is generally the case for the relatively sparse vegetation in a small chamber. However large chambers, which completely enclose a crop canopy or forest ground cover, are now being commonly used in trace-gas-emission studies from crops and natural ecosystems. (1989; Butterbach-Bahl et al., 1997). Even in well-stirred chambers, the plant canopy might constitute a major diffusive barrier, and therefore might introduce additional errors to flux estimation. We have investigated the influence on the measured trace gas flux of a deep canopy contained within a closed static chamber.

The concentration of a trace gas within a plant canopy can be described by the diffusion equation as follows:

$$\frac{\partial C}{\partial t} = \frac{\partial}{\partial z} \left( D \frac{\partial C}{\partial z} \right) = - \frac{\partial F}{\partial z} \quad (10)$$

where  $C$  is the gas concentration at a height  $z$  above the soil surface (m),  $t$  is time (s),  $D$  is the turbulent diffusion coefficient within the canopy ( $\text{m}^2 \text{s}^{-1}$ ),  $F$  is the volumetric vertical trace gas flux within the canopy ( $\text{m}^3 \text{m}^{-2} \text{s}^{-1}$ ) and  $z$  is the height above the soil surface (m). The following boundary conditions are applicable for this study:  $F = F_0$  when  $z = 0$ ;  $C = C_a$  when  $z = 0$ ;  $C = C_a$  when  $z = h$  (canopy height) and the chamber is open, where  $C_a$  is the ambient trace gas concentration;  $D = 0$  when  $z = h$  and chamber is closed.

Eq. 10 can be solved either analytically or numerically. The analytical solution can be formulated using the above boundary conditions for constant  $D$  (see p. 113, Carslaw and Jaeger, 1959). When  $D$  varies with  $z$  and the chamber is open, the steady state solution of Eq. 1 is given by

$$C(z) = C_a + F_0 \int_z^h \frac{dz}{D(z)}. \quad (11)$$

The term  $\int_z^h \frac{dz}{D(z)}$  is the total aerodynamic resistance between height  $z$  and  $h$ . Variation of  $D$  with  $z$  is calculated using the theory developed by Raupach (1987). A numerical solution is required for the time dependent solution after the chamber lid has closed. Information from this solution for the example of  $\text{N}_2\text{O}$  is provided in Fig. 4 and Fig. 5. The canopy height is 0.8 m. Turbulence is assumed to be patterned on a plane mixing layer rather than a surface boundary layer above the canopy, therefore the diffusivity is roughly constant.

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Address and contact details: CSIRO Atmospheric Research  
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